

## **Instructions for use**

**FlexStar<sup>®</sup>**

**LDT (RT-)PCR Detection Mix 1.5**

06/2023 EN



# FlexStar<sup>®</sup>

## LDT (RT-)PCR Detection Mix 1.5

For research use only!

(RUO)



FS0231503



96



06 2023



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## 1. Application

The FlexStar® LDT (RT-)PCR Detection Mix 1.5 is a buffered solution, which includes primers and a fluorescence labeled probe for the amplification and detection of the AltoStar® Internal Control 1.5. In combination with the FlexStar® (RT-)PCR Amplification Mix 1.5 the FlexStar® LDT (RT-)PCR Detection Mix 1.5 can be used for the real-time PCR based amplification and detection of nucleic acids.

**For research use only (RUO)! Not for use in diagnostic procedures.**

## 2. Product content

The FlexStar® LDT (RT-)PCR Detection Mix 1.5 contains the following components:

**Table 1:** Kit components

Lid color	Component	Number of tubes	Nominal volume [µl/tube]
Blue	Detection Mix <sup>1)</sup>	8	48
White	NTC <sup>2)</sup>	2	250

<sup>1)</sup> Contains biological material of animal origin

<sup>2)</sup> No Template Control (negative control)

## 3. Storage

- The FlexStar® LDT (RT-)PCR Detection Mix 1.5 is shipped on dry ice. The product components should arrive frozen. If one or more components are not frozen upon receipt, or if tubes have been compromised during shipment, contact Altona Diagnostics technical support for assistance (see chapter 9. Technical assistance).
- All components should be stored at -25 °C to -15 °C upon arrival.
- Repeated thawing and freezing of the Detection Mix component should be avoided, as this might affect the performance of the product.

- Repeated thawing and freezing of the No Template Control (NTC) (more than 4 times) should be avoided, as this might affect the performance of the product.
- Storage at room temperature (max. +30 °C) should not exceed a period of 2 hours.
- Protect the Detection Mix component from light.

### 4. Product description

The FlexStar® LDT (RT-)PCR Detection Mix 1.5 is a reagent system. Used in combination with the FlexStar® (RT-)PCR Amplification Mix 1.5 and target specific primers and probes it allows the amplification and detection of nucleic acid.

The FlexStar® LDT (RT-)PCR Detection Mix 1.5 is based on real-time (RT-) PCR technology, utilizing reverse-transcriptase (RT) reaction to convert RNA into complementary DNA (cDNA) and polymerase chain reaction (PCR) for the amplification of specific target sequences and target specific probes for the detection of the amplified DNA.

The FlexStar® LDT (RT-)PCR Detection Mix 1.5 includes oligonucleotides for the amplification and detection of the internal control (IC, AltoStar® Internal Control 1.5). The IC is automatically added at the beginning of the nucleic acid purification procedure on the AltoStar® Automation System AM16 (in the following summarized as AltoStar® AM16). For details refer to the instructions for use of the AltoStar® Internal Control 1.5.

The probe specific for the IC is labeled with the fluorophore JOE™. Primers and probes for the target specific amplification and detection have to be added.

Using probes linked to distinguishable dyes enables the parallel detection of specific target DNA/RNA and the IC in the corresponding detection channels of the real-time PCR instrument.

### 4.1 Components

The FlexStar® LDT (RT-)PCR Detection Mix 1.5 contains enough reagents for 96 reactions. The product consists of the following components:

- Detection Mix<sup>1)</sup>
- NTC<sup>2)</sup>

<sup>1)</sup> Contains biological material of animal origin

<sup>2)</sup> No Template Control (negative control)

Except for the DNA polymerase and the reverse transcriptase, which are included in the FlexStar® (RT-)PCR Amplification Mix 1.5, the Detection Mix component contains all reagents (PCR buffer, magnesium salt, primers and probes) to allow detection of the IC. Primers and probes for the detection of specific target DNA/RNA have to be added.

The NTC contains the IC template. The NTC is used as negative control for the target specific real-time PCR and indicates possible contamination of the Detection Mix component.

### 4.2 Real-time PCR instruments

The FlexStar® LDT (RT-)PCR Detection Mix 1.5 can be used with the following real-time PCR instruments:

- CFX96™ Deep Well Dx System (Bio-Rad)
- CFX96™ Dx System (Bio-Rad)
- LightCycler® 480 Instrument II (Roche)
- QuantStudio™ 5 Real-Time PCR System (Applied Biosystems)
- Rotor-Gene® Q5/6 plex Platform (QIAGEN)

#### NOTE



Ensure that all instruments used have been installed, calibrated, checked and maintained according to the manufacturer's instructions and recommendations.

## 5. Material required but not provided

The following additional instruments and consumables are required for use of the FlexStar® LDT (RT-)PCR Detection Mix 1.5 but not provided with this product:

- Appropriate real-time PCR instrument (see chapter 4.2 Real-time PCR instruments)
- Appropriate nucleic acid extraction system or kit (see chapter 6.1 Sample preparation)
- Vortex mixer
- Centrifuge (e.g. desktop centrifuge) for centrifugation of kit reagents
- Centrifuge for centrifugation of PCR plates
- Appropriate 96 well reaction plates or reaction tubes with corresponding (optical) closing material
- Pipettes (adjustable)
- Pipette tips with filters (disposable)
- Powder-free gloves (disposable)

Reagents required but not included in the FlexStar® LDT (RT-)PCR Detection Mix 1.5:

- FlexStar® (RT-)PCR Amplification Mix 1.5 (Order No. FS0011503/FS0011505)
- AltoStar® Internal Control 1.5 (Order No. IC15-06)
- Primer/probe mix for the specific nucleic acid amplification and detection (not supplied by altona Diagnostics)

## 6. Procedure

### 6.1 Sample preparation

Extracted nucleic acid is the starting material for the FlexStar® LDT (RT-)PCR Detection Mix 1.5. The quality of the extracted nucleic acid has a profound impact on the performance of the product.



For additional information and technical support regarding pre-treatment and sample preparation, contact altona Diagnostics technical support (see chapter 9. Technical assistance).

### 6.2 Preparation of the Detection Mix

Add 30 µl specific primer/probe mix (not supplied by altona Diagnostics) per tube of Detection Mix.

### 6.3 Master mix setup

All reagents and samples should be thawed completely, mixed (by pipetting or gentle vortexing) and centrifuged briefly before use.

The FlexStar® LDT (RT-)PCR Detection Mix 1.5 is configured for use with the FlexStar® (RT-)PCR Amplification Mix 1.5 and the AltoStar® Internal Control 1.5, which allows to control the sample preparation procedure (nucleic acid extraction) and the subsequent (RT-)PCR.

The FlexStar® LDT (RT-)PCR Detection Mix 1.5 contains primers and probes for the AltoStar® Internal Control 1.5 (IC), which can either be used as (RT-)PCR inhibition control or as a control of the sample preparation procedure (nucleic acid extraction) and as a (RT-)PCR inhibition control.

- ▶ The IC is automatically added at the beginning of the nucleic acid purification procedure on the AltoStar® AM16.
- ▶ When using other nucleic acid extraction methods, the IC has to be added during the lysis step either manually or automatically by the respective instrument.
- ▶ No matter which method/system is used for nucleic acid extraction, never add the IC directly to the sample. The IC should always be added to the sample/lysis buffer mixture. The volume of the IC which has to be added, always and only depends on the elution volume. It represents 50 % of the elution volume. For instance, if the nucleic acid is going to be eluted in 60 µl of elution buffer or water, 30 µl of IC per sample must be added into the sample/lysis buffer mixture.

- ▶ Set up the master mix according to the following pipetting scheme:

**Table 2:** Pipetting scheme (master mix setup)

Number of reactions (rxns)	1	12
Detection Mix (incl. specific primers/probes)	5 µl*	60 µl
Amplification Mix	15 µl	180 µl
<b>Volume master mix</b>	<b>20 µl</b>	<b>240 µl</b>

\* Composed of 1 µl specific primers/probes and 4 µl Detection Mix

#### NOTE



No matter which method/system is used for nucleic acid extraction, never add the IC directly to the specimen.

## 6.4 Reaction setup

- ▶ Pipette 20 µl of the master mix into each required well of an appropriate optical 96-well reaction plate or an appropriate optical reaction tube.
- ▶ Add 10 µl of the sample (eluate from the nucleic acid extraction) or 10 µl of the controls (specific positive control or NTC).

**Table 3:** Pipetting scheme (reaction setup)

Reaction setup	
Master mix	20 µl
Sample or control	10 µl
<b>Total volume</b>	<b>30 µl</b>

- ▶ Make sure that at least 1 specific positive control (not supplied by Altona Diagnostics) and 1 NTC is used per run.

- ▶ Thoroughly mix the samples and controls with the master mix by pipetting up and down.
- ▶ Close the 96-well reaction plate with appropriate lids or optical adhesive film and the reaction tubes with appropriate lids.
- ▶ Centrifuge the 96-well reaction plate in a centrifuge with a microtiter plate rotor and the reaction tubes in an appropriate centrifuge for 30 seconds at approximately 1,000 x g (~ 3,000 rpm).
- ▶ The NTC does already contain the IC template in the correct concentration.

## 7. Programming the real-time PCR instrument

For basic information regarding the setup and programming of the different real-time PCR instruments, refer to the user manual of the respective instrument.

For detailed programming instructions regarding the use of the FlexStar® LDT (RT-)PCR Detection Mix 1.5 on specific real-time PCR instruments, contact altona Diagnostics technical support (see chapter 9. Technical assistance).

### 7.1 Settings

- ▶ Define the following settings:

**Table 4:** Run settings

Settings	
Reaction volume	30 µl
Ramp rate	Default
Passive reference*	None

\* If applicable

## 7.2 Fluorescence detectors (dyes)

- Define the fluorescence detectors (dyes):

**Table 5:** Fluorescence detectors

Target	Detector name	Reporter	Quencher
Specific nucleic acid	Depends on the specific primer/probe mix		
IC	Internal Control	JOE™	(None)

## 7.3 Temperature profile and dye acquisition

- Define the temperature profile and dye acquisition:

**Table 6:** Temperature profile and dye acquisition

	Stage	Cycle repeats	Acquisition	Temperature [°C]	Time [min:s]
Reverse transcription	Hold	1	-	52	05:00
Denaturation	Hold	1	-	95	00:05
Amplification	Cycling	45	-	95	00:05
			Yes	58	00:25

## 8. Data analysis

For basic information regarding data analysis on specific real-time PCR instruments, refer to the user manual of the respective instrument.

For detailed instructions regarding the analysis of the data generated with the FlexStar® LDT (RT-)PCR Detection Mix 1.5 on different real-time PCR instruments, contact altona Diagnostics technical support (see chapter 9. Technical assistance).

## 8.1 Interpretation of results

The Internal Control serves as (RT-)PCR inhibition control or as a control of the sample preparation procedure (nucleic acid extraction) and as a (RT-)PCR inhibition control. Detection of the IC in the JOE™ detection channel is not required for positive results in the specific detection channel. A high target specific RNA/DNA load in the sample can lead to reduced or absent IC signals.

## 9. Technical assistance

For customer support, contact altona Diagnostics technical support:

**e-mail:**                **support@altona-diagnostics.com**  
**phone:**                **+49-(0)40-5480676-0**

## 10. Trademarks and disclaimers














AltoStar®, FlexStar® (altona Diagnostics); QuantStudio™ (Applied Biosystems); CFX96™ (Bio-Rad); Rotor-Gene® (QIAGEN); LightCycler® (Roche); FAM™, JOE™ (Thermo Fisher Scientific).



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## 11. Explanation of symbols

Symbol	Explanation
	Research use only
	Batch code
	Content
	Cap color
	Catalogue number
	Number
	Component
	Consult instructions for use
	Contains sufficient for "n" tests/reactions (rxns)
	Temperature limit
	Use-by date
	Manufacturer
	Material number

Symbol	Explanation
	Version
	Note: Information is given to the user that is useful but not essential to the task at hand.

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