

Instructions for use

ExtraStar® Purification Kit 2.0

12/2022 EN

ExtraStar® Purification Kit 2.0

For use with

KingFisher™ Flex Purification System (Thermo Fisher Scientific)

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IVD

REF 5012045

<u>Σ</u> 384

12 2022

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ExtraStar® Purification Kit 2.0

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1. About these instructions for use

These instructions for use guide the user in utilizing the ExtraStar® Purification Kit 2.0 in combination with the AltoStar® Internal Control 1.5 on the KingFisher™ Flex Purification System (Thermo Fisher Scientific).

The main operation steps of the KingFisher™ Flex Purification System and the AltoStar® Internal Control 1.5 during the purification procedure are described for comprehensibility.

For more detailed information about these products, refer to the respective instructions for use listed below:

- KingFisher™ Flex Purification System Manual (Thermo Fisher Scientific)
- Instructions for use AltoStar® Internal Control 1.5

Throughout this manual, the terms CAUTION and NOTE have the following meanings:

CAUTION



Highlights operating instructions or procedures which, if not followed correctly, may result in personal injury or impact product performance. Contact altona Diagnostics technical support for assistance.

NOTE



Information is given to the user that is useful but not essential to the task at hand.

Read the instructions for use carefully before using the product.

2. Intended use

The ExtraStar® Purification Kit 2.0 uses magnetic particle technology and is intended to be used for the automated isolation and purification of nucleic acids from human respiratory swab specimens for *in vitro* diagnostic purposes.

The product is designed for use with the KingFisher[™] Flex Purification System (Thermo Fisher Scientific) and altona Diagnostics kits and reagents specified for use with the ExtraStar[®] Purification Kit 2.0.

The ExtraStar® Purification Kit 2.0 is intended for use by professional users trained in molecular biological techniques and *in vitro* diagnostic procedures.

3. Kit content

The ExtraStar® Purification Kit 2.0 is shipped in 2 separate boxes **Box 1** and **Box 2** (see tables 1 and 2).

Table 1: Kit components Box 1

Component	Number of bottles	Volume per bottle [ml]
Lysis Buffer	2	120
Wash Buffer 1	2	100
Wash Buffer 2	2	100
Wash Buffer 3	2	100

Table 2: Kit components Box 2

Component	Number of bottles	Volume per bottle [ml]
Magnetic Beads	2	5
Elution Buffer	2	22
Enhancer	2	4



Before first use check the product and its components for completeness with respect to number, type and filling. Do not use a defective or incomplete product, as product performance could be compromised.

The ExtraStar® Purification Kit 2.0 contains reagents sufficient for 384 sample purifications.

Upon receipt check the product and its components for:

- Integrity
- Completeness with respect to number, type and filling
- Correct labeling
- Expiration date
- Clarity and absence of particles

If any kit component has been compromised during shipment or is missing, contact altona Diagnostics technical support for assistance (see chapter 14. Technical support).

4. Storage and handling

All reagents included in the ExtraStar® Purification Kit 2.0 are ready-to-use solutions.

4.1 Storage

The ExtraStar® Purification Kit 2.0 is shipped at room temperature. **Box 1** has to be stored at +15 °C to +30 °C and **Box 2** has to be stored at +2 °C to +8 °C upon receipt (see table 3). The bottles must be stored in an upright position.

Table 3: Storage conditions for Box 1 and Box 2

Storage conditions			
Box 1 Box 2			
+15 °C to +30 °C	+2 °C to +8 °C		

CALITION



Improper storage conditions could compromise product performance.

CAUTION



Do not use products beyond the expiration date. The use of expired products could compromise product performance.

4.2 Handling

The reagents of the ExtraStar® Purification Kit 2.0 are stable after initial opening for 14 days, when closed after each use and stored as follows: Magnetic Beads, Enhancer and Elution Buffer shall be closed with the original cap after use and stored at +2 °C to +8 °C. The Lysis Buffer and Wash Buffer 1, 2 and 3 shall be closed with the original cap after use and stored at +15 °C to +30 °C.

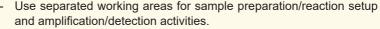
CAUTION



Do not leave reagents open in between use, as this could compromise product performance.

Improper handling of product components and samples may cause contamination and could compromise product performance:

- Do not interchange bottle caps.
- Store positive and/or potentially positive material separated from the kit components.



- Always dispose gloves after handling positive and/or potentially positive material.
- Do not open the PCR plates and/or tubes post amplification.

CAUTION



Do not exceed handling durations as specified in these instructions for use, as this could compromise product performance.

CAUTION



Do not mix components from different kit lots, as this could compromise product performance.

NOTE



The Lysis Buffer may crystalize at low temperature. If crystals occur, the Lysis Buffer bottle should be heated with careful pivoting (\leq +50 °C, e.g., in a water bath) until the crystals are completely dissolved (up to 30 min).

NOTE



The Magnetic Beads should be shaken well before use (e.g., vortex for 60 seconds).

NOTE



Slight colour changes may occur to the Lysis Buffer. These slight changes in colour do not indicate a change in the quality of the buffer.

5. Product description

Table 4: Components of the ExtraStar® Purification Kit 2.0

Kit component	Description
Lysis Buffer	The Lysis Buffer contains chaotropic salts and surfactants (guanidine thiocyanate, octoxynol) to disrupt cells or virions chemically. It stabilizes nucleic acids and protects them against nucleases in solution.
Wash Buffer 1	Wash Buffer 1 contains different salts and organic solvents (guanidine thiocyanate and ethanol) to remove proteins and other impurities.
Wash Buffer 2	Wash Buffer 2 contains organic solvents (ethanol) to remove proteins and other impurities.
Wash Buffer 3	Wash Buffer 3 contains different salts in order to purify the nucleic acids.
Enhancer	The Enhancer stabilizes and protects nucleic acids against nucleases in solution.
Magnetic Beads	The Magnetic Beads are coated with a thin layer of silica to bind free nucleic acids in solution. The magnetic characteristic allows the separation of beads from liquids in a magnetic field.
Elution Buffer	The Elution Buffer is a low-salt buffer to release the nucleic acids from the Magnetic Beads for subsequent analysis.

5.1 Principle of method

The ExtraStar® Purification Kit 2.0 is intended for the automated isolation and purification of nucleic acids from specified human specimens (see chapter 6. Sample types) for *in vitro* diagnostic purposes in conjunction with the KingFisher™ Flex Purification System, the AltoStar® Internal Control 1.5 and altona Diagnostics kits and reagents specified for use with the ExtraStar® Purification Kit 2.0. The ExtraStar® Purification Kit 2.0 is based on magnetic bead technology, utilizing silica coated magnetic particles, which can bind and release nucleic acids under specific conditions [1,2,3].

The purification procedure comprises 3 automated steps on the KingFisher™ Flex Purification System (see figure 1).

- In the first step nucleic acids are released by chemical and mechanical lysis under chaotropic high salt conditions. The conditions stabilize the nucleic acids in solution and enable their binding to the magnetic silica beads.
- In the following washing steps different wash buffers are used to remove proteins and other impurities.
- 3. Finally, the nucleic acids are released from the magnetic beads with an elution buffer into the eluate plate.

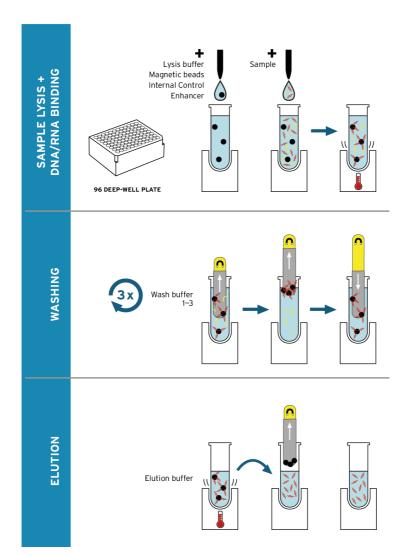


Figure 1: Illustration of the purification procedure using the KingFisher™ Flex Purification System

6. Sample types

The following sample type is validated for use with the ExtraStar® Purification Kit 2.0:

· Human respiratory swabs in transport medium

CAUTION



Do not use other sample types! The use of other sample types could compromise product performance.

NOTE



Frozen storage of samples does not compromise kit performance. When working with frozen samples, make sure samples are completely thawed and properly mixed before use.

NOTE



For information regarding collection, handling and storage of samples refer to the instructions for use of altona Diagnostics kits and reagents specified for use with the ExtraStar® Purification Kit 2.0.

7. Warnings, precautions and limitations

	Lysis Buffer				
	H302+H312+H332	Harmful in contact with skin or if inhaled or swallowed.			
(工業)	H314	Causes severe skin burns and eye damage.			
	H411	Toxic to aquatic life with long lasting effects.			
GHS05	EUH032	Contact with acids liberates very toxic gas.			
	EUH071	Corrosive to the respiratory tract.			
	P260	Do not breathe mist, vapours, spray.			
(1)	P264	Wash hands thoroughly after handling.			
GHS07	P273	Avoid release to the environment.			
GHS07	P280	Wear protective clothing, eye protection, face protection, protective gloves.			
¥2>	P303+P361+P353	IF ON SKIN (or hair): Take off immediately all contaminated clothing. Rinse skin with water or shower.			
	P310	Immediately call a POISON CENTER, a doctor.			
GHS09	Contains:	Guanidine thiocyanate (CAS 593-84-0) 50-70 %.			
Danger!		Alkylphenol ethoxylat (CAS 9036-19-5) 10-20 %.			

	H226	Flammable liquid and vapour.
(4)	H314	Causes severe skin burns and eye damage.
	H412	Harmful to aquatic life with long lasting effect.
GHS02	EUH032	Contact with acids liberates very toxic gas.
	EUH071	Corrosive to the respiratory tract.
I B	P210	Keep away from heat, hot surfaces, sparks, open flames and other ignition sources. No smoking.
	P260	Do not breathe mist, vapours, spray.
GHS05	P273	Avoid release to the environment.
anger!	P280	Wear protective clothing, eye protection, face protection, protective gloves.
	P303+P361+P353	IF ON SKIN (or hair): Take off immediately all contaminated clothing. Rinse skin with water or shower.
	P310	Immediately call a POISON CENTER, a doctor.
	Contains:	Guanidine thiocyanate (CAS 593-84-0) 25–50 %.
		Ethanol (CAS 64-17-5) 25-50 %.
		Wash Buffer 2
	H226	Flammable liquid and vapour.
30%	H319	Causes serious eve irritation

	H226	Flammable liquid and vapour.		
<u> </u>	H319	Causes serious eye irritation.		
GHS02	P210	Keep away from heat, hot surfaces, sparks, open flames and other ignition sources. No smoking.		
	P233	Keep container tightly closed.		
	P280	Wear protective clothing, eye protection, face protection, protective gloves.		
GHS07	P305+P351+P338	IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.		
Danasal	P337+P313	If eye irritation persists: Get medical advice/attention.		
Danger!	Contains:	Ethanol (CAS 64-17-5) 50-70 %.		

Enhancer			
	H314	Causes severe skin burns and eye damage.	
不多	P260	Do not breathe mist, vapours, spray.	
GHS05	P280	Wear protective clothing, eye protection, face protection, protective gloves.	
Danger!	P303+P361+P353	IF ON SKIN (or hair): Take off immediately all contaminated clothing. Rinse skin with water or shower.	
	P305+P351+P338	IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.	
	P310	Immediately call a POISON CENTER, a doctor.	
	Contains:	Tris(2-carboxyethyl)phosphine (CAS 51805-45-9) 10–20 %.	

NOTE



For more information, consult the safety data sheets (SDSs).

- Before first use check the product and its components for completeness with respect to number, type and filling. Do not use a defective or incomplete product, as product performance could be compromised.
- Improper storage conditions could compromise product performance.
- Do not use products beyond the expiration date. The use of expired products could compromise product performance.
- Do not leave reagents open in between use, as this could compromise product performance.
- Improper handling of product components and samples may cause contamination and could compromise product performance:
 - Do not interchange bottle caps.
 - Store positive and/or potentially positive material separated from the kit components.
 - Use separated working areas for sample preparation/reaction setup and amplification/detection activities.
 - Always dispose gloves after handling positive and/or potentially positive material.
 - Do not open the PCR plates and/or tubes post amplification.
- Do not exceed handling durations as specified in these instructions for use, as this could compromise product performance.
- Do not mix components from different kit lots, as this could compromise product performance.
- Do not use other sample types! The use of other sample types could compromise product performance.
- Do not fill more volume into plate wells as specified, as this could lead to cross-contamination and could compromise product performance.
- Always use the correct sample volume when preparing the lysis sample plate, otherwise the product performance could be compromised.

- Always fill the correct buffer into the corresponding buffer plate. Mix up of buffers could compromise product performance.
- Always check for enough buffers for each sample before starting the experiment. The use of less buffer volume than specified could compromise product performance.
- Always use the correct KingFisher™ programming for extraction process, as other settings could lead to cross-contamination and could compromise product performance.
- Make sure to fill the corresponding well positions on each plate. Do not mix up sample and buffer positions on the plate wells as this could compromise product performance.
- Do not mix up well plates and plate orientation while loading the KingFisher™.
 Incorrect loading of plates could compromise product performance.
- Calcium alginate swabs, swabs with wooden shafts and/or cotton tips as well as swabs containing jelly agar may reduce extraction performance.
- Improper preparation of reagents (e.g., Lysis Buffer and Magnetic Beads) may cause invalid or false negative results.
- Do not interchange bottle caps when closing product components after use to avoid contamination of reagents, which could compromise product performance.
- Do not use samples which contain solids and high-viscosity constituents, as this could compromise product performance.
- Storage of eluates under wrong conditions may lead to loss of eluate volume and/or degradation of the pathogen specific target sequence and could compromise product performance.
- Always treat samples as infectious and (bio-)hazardous material in accordance with safety and laboratory procedures. For sample material spills promptly use an appropriate disinfectant. Handle contaminated materials as biohazardous.
- Disposal of hazardous and biological waste shall comply with local and national regulations to avoid environmental contamination.

8. Using the ExtraStar® Purification Kit 2.0

The following chapters describe the use of the ExtraStar® Purification Kit 2.0.

8.1 Sample volume

The ExtraStar® Purification Kit 2.0 allows purification of 300 µl sample.

CAUTION



Do not fill more volume into plate wells as specified, as this could lead to cross-contamination and could compromise product performance.

CAUTION



Always use the correct sample volume when preparing the lysis sample plate, otherwise the product performance could be compromised.

8.2 Material and devices required but not provided

- Nucleic acid extraction and PCR amplification and detection control AltoStar[®] Internal Control 1.5 (altona Diagnostics Ordner No. IC15-46)
- Thermo Fisher Scientific KingFisher[™] 96 Flex with 96 deep-well magnet and heating block with Thermo Fisher Scientific BindIt[™] 4.0 Software or higher (Thermo Fisher Scientific Order No. 5400630)
- 4x KingFisher™ 96 deep-well plate (Thermo Fisher Scientific Order No. 95040450)
- 1x KingFisher™ 96 tip comb for deep-well magnets (Thermo Fisher Scientific Order No. 97002534)
- 2x KingFisher™ 96 plate 200 µl (Thermo Fisher Scientific Order No. 97002540)

NOTE



Ensure that all instruments used have been installed, calibrated, checked and maintained according to the manufacturer's instructions and recommendations.

8.3 General material and devices

- Vortex mixer
- Powder-free gloves (disposable)
- Pipettes (adjustable, for reagent and sample preparation)
- Pipette tips with filters (disposable, for sample preparation)
- Optional: stepper pipette (adjustable, for reagent preparation) and suitable tips (disposable)

9. Purification using the ExtraStar® Purification Kit 2.0 in combination with the KingFisher™ Flex

For first use, a method specified for the sample type must be created using Thermo Scientific™ BindIt™ 4.0 Software or higher for KingFisher™ instruments. This method can be used on the instrument as a stand-alone unit or with a connected PC. For use of BindIt™ Software and programming, refer to the relevant user manual of the device.

Table 5: KingFisher™ program for ExtraStar® Purification Kit 2.0 extraction

KingFisher™ program for ExtraStar® Purification Kit 2.0 extraction				
Lysis samp	ole plate	96 deep-well plate		
Name Well volume [µl]		Total reagent volume [μl]	Туре	
Lysis Buffer	500	-	Reagent	
Magnetic Beads	25	-	Reagent	
Enhancer	20	-	Reagent	
AltoStar [®] Internal Control 1.5	50	-	Reagent	
Sample	300	-	Sample	

KingFisher™ program for ExtraStar® Purification Kit 2.0 extraction				
Wash 1 plate		96 deep-well plate		
Name	Well volume [µl]	Total reagent volume [µl]	Туре	
Wash Buffer 1	500	-	Reagent	
Wash 2	plate	96 deep-	well plate	
Name	Well volume [µl]	Total reagent volume [μl]	Туре	
Wash Buffer 2	500	-	Reagent	
Wash 3	plate	96 deep-well plate		
Name	Well volume [µl]	Total reagent volume [μl]	Туре	
Wash Buffer 3	500	- Reagen		
Eluate _l	plate	96 standard plate		
Name	Well volume [µl]	Total reagent volume [µl]	Туре	
Elution Buffer	100	-	Reagent	
Comb plate		96 standard plate		
Name	Well volume [µl]	Total reagent Type volume [µl]		
-	-	-	-	

CALITION



Always fill the correct buffer into the corresponding buffer plate. Mix up of buffers could compromise product performance.

CAUTION



Always check for enough buffers for each sample before starting the experiment. The use of less buffer volume than specified could compromise product performance.



Do not fill more volume into plate wells as specified, as this could lead to cross-contamination and could compromise product performance.

Table 6: Protocol

KingFisher™ protocol for ExtraStar® Purification Kit 2.0 extraction			
Tip 1	96 deep-well tip comb		
Pick-up	Comb plate		
Lysis	Lysis sample plate		
Beginning of step	Precollect	No	
Degining of step	Release time, speed	No	
	Shake 1 time, speed	00:00:30, slow	
	Shake 2 time, speed	00:00:10, bottom mix	
Mixing/heating	Loop count	10	
Wiking/neating	Heating temperature [°C]	56	
	Preheat	Yes	
	Heating during mixing	Yes	
	Postmix	No	
End of step	Collect count	4	
	Collect time [s]	1	
Wash 1	Wash	1 plate	
Beginning of step	Precollect	No	
beginning of step	Release time, speed	00:00:10, slow	
	Shake 1 time, speed	00:00:30, slow	
Mixing/hooting	Shake 2 time, speed	00:00:10, bottom mix	
Mixing/heating	Loop count	4	
	Heating during mixing	No	

	Postmix	No	
End of step	Collect count [s]	3	
	Collect time [s]	0	
Wash 2	Wash 2 plate		
Beginning of step	Precollect	No	
beginning of step	Release time, speed	00:00:05, fast	
	Shake 1 time, speed	00:00:05, slow	
Mixing/heating	Shake 2 time, speed	00:00:30, bottom mix	
wiixii ig/neating	Loop count	3	
	Heating during mixing	No	
	Postmix	No	
End of step	Collect count	3	
	Collect time [s]	0	
Wash 3	Wash	3 plate	
	Wash	3 plate No	
Wash 3 Beginning of step			
	Precollect	No	
Beginning of step	Precollect Release time, speed	No 00:00:05, fast	
	Precollect Release time, speed Shake 1 time, speed	No 00:00:05, fast 00:00:05, slow	
Beginning of step	Precollect Release time, speed Shake 1 time, speed Shake 2 time, speed	No 00:00:05, fast 00:00:05, slow 00:00:30, bottom mix	
Beginning of step	Precollect Release time, speed Shake 1 time, speed Shake 2 time, speed Loop count	No 00:00:05, fast 00:00:05, slow 00:00:30, bottom mix 2	
Beginning of step	Precollect Release time, speed Shake 1 time, speed Shake 2 time, speed Loop count Heating during mixing	No 00:00:05, fast 00:00:05, slow 00:00:30, bottom mix 2 No	
Beginning of step Mixing/heating	Precollect Release time, speed Shake 1 time, speed Shake 2 time, speed Loop count Heating during mixing Postmix	No 00:00:05, fast 00:00:05, slow 00:00:30, bottom mix 2 No No	
Beginning of step Mixing/heating	Precollect Release time, speed Shake 1 time, speed Shake 2 time, speed Loop count Heating during mixing Postmix Collect count	No 00:00:05, fast 00:00:05, slow 00:00:30, bottom mix 2 No No 3 0	
Beginning of step Mixing/heating End of step	Precollect Release time, speed Shake 1 time, speed Shake 2 time, speed Loop count Heating during mixing Postmix Collect count Collect time [s]	No 00:00:05, fast 00:00:05, slow 00:00:30, bottom mix 2 No No 3 0	

Missionelleanting	Mixing time, speed	00:10:00, slow
	Heating temperature [°C]	70
Mixing/heating	Preheat	No
	Heating during mixing Yes	
End of step	Postmix	No
	Collect count	3
	Collect time [s] 0	
Leave	Wash 2 plate	



Always use the correct KingFisher™ programming for extraction process, as other settings could lead to cross-contamination and could compromise product performance.

9.1 Transport medium (swab rinse)

- Make sure that the method (see table 5) is programmed and installed on the KingFisher™ Flex.
- 2. Prepare an empty standard plate (KingFisher™ 96 plate 200 µl) with a 96 deep-well tip comb.
- 3. Prepare the wash 1 plate (KingFisher™ 96 deep-well plate) by adding 500 µl Wash Buffer 1 into each well in use.
- 4. Prepare the wash 2 plate (KingFisher™ 96 deep-well plate) by adding 500 µl Wash Buffer 2 into each well in use.
- Prepare the wash 3 plate (KingFisher™ 96 deep-well plate) by adding 500 µl
 Wash Buffer 3 into each well in use.
- Prepare the eluate plate (KingFisher[™] plate 200 µl) by adding 100 µl Elution Buffer into each well in use.
- 7. Sample preparation for transport medium (swab rinse):

Prepare the Lysis sample plate (KingFisher $^{\text{TM}}$ 96 deep-well plate) by adding in following order into each well in use:

500 µl Lysis Buffer

- 25 µl of well mixed (e.g., 60 seconds shaked or votexed) Magnetic Beads
- 20 µl of Enhancer solution
- 300 µl of sample (e.g., viral transport medium)
- **8.** Start the extraction method immediately and follow the instructions by placing the plates into the instrument.
- 9. Start the run (it takes approximately 30 min to finish).
- **10.** After finishing the run use the eluate plate for PCR process.



Always fill the correct buffer into the corresponding buffer plate. Mix up of buffers could compromise product performance.

CAUTION



Always check for enough buffers for each sample before starting the experiment. The use of less buffer volume than specified could compromise product performance.

CAUTION



Do not fill more volume into plate wells as specified, as this could lead to cross-contamination and could compromise product performance.

CAUTION



Make sure to fill the corresponding well positions on each plate. Do not mix up sample and buffer positions on the plate wells as this could compromise product performance.

CAUTION



Do not mix up well plates and plate orientation while loading the KingFisher $^{\text{TM}}$. Incorrect loading of plates could compromise product performance.



Calcium alginate swabs, swabs with wooden shafts and/or cotton tips as well as swabs containing jelly agar may reduce extraction performance.

CAUTION



Improper preparation of reagents (e.g., Lysis Buffer and Magnetic Beads) may cause invalid or false negative results.

CAUTION



Do not interchange bottle caps when closing product components after use to avoid contamination of reagents, which could compromise product performance.

CAUTION



Do not use samples which contain solids and high-viscosity constituents, as this could compromise product performance.

CAUTION



Always use the correct sample volume when preparing the lysis sample plate, otherwise the product performance could be compromised.

9.1.1 Eluate stability

After completion of the purification run the eluates in the unsealed eluate plate are stable at room temperature (max. +30 °C) for a total of 4 hours.

CAUTION



Storage of eluates under wrong conditions may lead to loss of eluate volume and/or degradation of the pathogen specific target sequence and could compromise product performance.

10. Performance data

The performance of the ExtraStar® Purification Kit 2.0 is verified in conjunction with each altona Diagnostics real-time PCR kit or reagent specified for use with the ExtraStar® Purification Kit 2.0. For information on performance data, refer to the instructions for use of the respective altona Diagnostics real-time PCR kit or reagent.

11. Disposal

Dispose of hazardous and biological waste in compliance with local and national regulations. Leftover product components and waste should not be allowed to enter sewage, water courses or soil.

CAUTION



Always treat samples as infectious and (bio-)hazardous material in accordance with safety and laboratory procedures. For sample material spills promptly use an appropriate disinfectant. Handle contaminated materials as biohazardous.

CAUTION



Disposal of hazardous and biological waste shall comply with local and national regulations to avoid environmental contamination.

NOTE



Liquid waste and any liquids containing Lysis Buffer or Wash Buffer 1 contain guanidine thiocyanate, which can form toxic, highly reactive and volatile compounds when combined with bleach or strong acids.

12. Quality control

In accordance with the altona Diagnostics GmbH EN ISO 13485-certified Quality Management System, each lot of ExtraStar® Purification Kit 2.0 is tested against predetermined specifications to ensure consistent product quality.

13. Troubleshooting guide

Problem: precipitate in reagent

Possible cause	Suggestions
Storage of the Lysis Buffer at low temperature or prolonged storage	If the Lysis Buffer bottle is already opened, make sure to close it with the same lid. Heat the Lysis Buffer bottle (≤ +50 °C, e.g., in a water bath) with careful intermittent pivoting until the precipitates are completely dissolved.
Excessive evaporation due to improper use and/or closing may lead to increased salt concentration in reagents	Discard the reagent. Make sure to always close the reagent bottles immediately after use.

Problem: low yield or purity of nucleic acids

Possible cause	Suggestions
Storage of reagents under wrong conditions	Discard reagents. Make sure to store the product components under defined storage conditions (see chapter 4. Storage and handling).
Reagents were not closed and/or stored properly in between use	Discard reagents. Make sure to store the product components under defined storage conditions (see chapter 4. Storage and handling). Make sure to always close the reagent bottles immediately after use.
Improper preparation of samples	Make sure to prepare samples according to the instructions in chapter 9.1 Transport medium (swab rinse).
Frozen samples were not thawed or mixed properly	Make sure samples are completely thawed and properly mixed before use.
Incomplete sample lysis	Before use, check that the Lysis Buffer does not contain precipitates. If the Lysis Buffer bottle is already opened, make sure to close the bottle with the corresponding lid and heat the bottle (≤ +50 °C, e.g., in a water bath) with careful intermittent pivoting until the precipitates are completely dissolved.

Possible cause	Suggestions
Mix up of buffers while filling the plates or mix up of buffer plates while loading the KingFisher™	Make sure to fill the correct buffers into the corresponding plates and load the plates according to the instructions of the method shown in KingFisher™ display.
High sample viscosity or solids in the sample	Make sure to prepare samples according to chapter 9.1 Transport medium (swab rinse).

14. Technical support

For customer support, contact altona Diagnostics technical support:

e-mail: support@altona-diagnostics.com

phone: +49-(0)40-5480676-0

NOTE



Any serious incident that has occurred in relation to this product shall be reported to altona Diagnostics and the competent authority of your country.

15. Literature

- [1] Mark A. Lever, Andrea Torti, Philip Eickenbusch, Alexander B. Michaud, Tina Šantl-Temkiv, and Bo Barker Jørgensen: A modular method for the extraction of DNA and RNA, and the separation of DNA pools from diverse environmental sample types; Front Microbiol. 2015; 6: 476.
- [2] Sonja Berensmeier: Magnetic particles for the separation and purification of nucleic acids; Appl Microbiol Biotechnol 2006 73:495–504.
- [3] Peter E. Vandeventer, Jessica S. Lin, Theodore J. Zwang, Ali Nadim, Malkiat S. Johal, and Angelika Niemz: Multiphasic DNAAdsorption to Silica Surfaces under Varying Buffer, pH, and Ionic Strength Conditions; J Phys Chem B. 2012 May 17; 116(19): 5661–5670.

16. Trademarks and disclaimers

AltoStar®, ExtraStar®, RealStar® (altona Diagnostics GmbH); BindIt™, KingFisher™ (Thermo Fisher Scientific).

Registered names, trademarks, etc. used in this document, even if not specifically marked as such, are not to be considered unprotected by law.

The ExtraStar® Purification Kit 2.0 is a CE-marked product according to the European *in vitro* diagnostic Regulation (EU) 2017/746.

Product not licensed with Health Canada and not FDA cleared or approved.

Not available in all countries.

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17. Symbols

Symbol	Explanation	
IVD	In vitro diagnostic medical device	
GTIN	Global Trade Item Number	
LOT	Batch code	
CONT	Content	
REF	Catalogue number	
NUM	Number	
СОМР	Component	
[]i	Consult instructions for use	
$\overline{\Sigma}$	Contains sufficient for "n" tests/reactions (rxns)	
A	Temperature limit	
	Use-by date	
	Manufacturer	
\triangle	Caution	

Symbol	Explanation	
MAT	Material number	
	Version	
i	Note	
UFI	Unique formula identifier	

18. Revision history

Table 7: Revision history

Identifier	Date of issue [month/year]	Modifications
MAN-5012040- EN-S01	12/2022	Initial release

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