

Instructions for Use

RealStar[®] JCV PCR Kit 1.2

02/2018 EN

Important!
Notice of change

Notice of Change

Please note that this assay reports results in
International Units (IU).

**For any further questions,
please contact us at**

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RealStar[®]

JCV PCR Kit 1.2

For research use only!

(RUO)



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1. Application

The RealStar® JCV PCR Kit 1.2 is a reagent system, based on real-time PCR technology, for the detection and quantification of JC virus (JCV) specific DNA.

For research use only (RUO)! Not for use in diagnostic procedures.

2. Kit Components

Lid Color	Component	Number of Vials	Volume [µl/Vial]
Blue	Master A	4	60
Purple	Master B	4	120
Green	Internal Control	1	1000
Red	QS1-4*	4	250
White	Water (PCR grade)	1	500

* The RealStar® JCV PCR Kit 1.2 contains Quantification Standards (QS) at four different concentrations (see Chapter 4. Product Description)

3. Storage

- The RealStar® JCV PCR Kit 1.2 is shipped on dry ice. The components of the kit should arrive frozen. If one or more components are not frozen upon receipt, or if tubes have been compromised during shipment, contact Altona Diagnostics GmbH for assistance.
- All components should be stored between -25°C and -15°C upon arrival.
- Repeated thawing and freezing of Master reagents (more than twice) should be avoided, as this might affect the performance of the assay. The reagents should be frozen in aliquots, if they are to be used intermittently.
- Storage between +2°C and +8°C should not exceed a period of two hours.
- Protect Master A and Master B from light.

4. Product Description

The RealStar® JCV PCR Kit 1.2 is a reagent system, based on real-time PCR technology, for the detection and quantification of JC virus (JCV) specific DNA.

The assay includes a heterologous amplification system (Internal Control) to identify possible PCR inhibition and to confirm the integrity of the reagents of the kit.

Real-time PCR technology utilizes polymerase chain reaction (PCR) for the amplification of specific target sequences and target specific probes for the detection of the amplified DNA. The probes are labelled with fluorescent reporter and quencher dyes.

Probes specific for JCV DNA are labelled with the fluorophore FAM™. The probe specific for the Internal Control (IC) is labelled with a fluorophore showing similar characteristics to Cy®3.

Using probes linked to distinguishable dyes enables the parallel detection of JCV specific DNA and the Internal Control in corresponding detector channels of the real-time PCR instrument.

The test consists of two processes in a single tube assay:

- PCR amplification of target DNA and Internal Control
- Simultaneous detection of PCR amplicons by fluorescent dye labelled probes

The RealStar® JCV PCR Kit 1.2 consists of:

- Two Master reagents (Master A and Master B)
- Internal Control (IC)
- Four Quantification Standards (QS1 - QS4)
- PCR grade water

Master A and Master B contain all components (PCR buffer, DNA polymerase, magnesium salt, primers and probes) to allow PCR mediated amplification and detection of JCV specific DNA and Internal Control in one reaction setup.

The Quantification Standards contain standardized concentrations of JCV specific DNA. These Quantification Standards were calibrated against the 1st World Health Organization International Standard for JC Virus for Nucleic Acid Amplification Techniques (NAT) (NIBSC code 14/114). The Quantification Standards can be used individually as positive controls, or together to generate a **standard curve**, which can be used to determine the concentration of JCV specific DNA in a sample.

The Quantification Standards have the following concentrations:

Quantification Standard	Concentration [IU/μl]
QS1	1.00E+04
QS2	1.00E+03
QS3	1.00E+02
QS4	1.00E+01

4.1 Real-Time PCR Instruments

The RealStar® JCV PCR Kit 1.2 can be used with the following real-time PCR instruments:

- SmartCycler® II (Cepheid)
- LightCycler® 1.2/1.5/2.0 Instruments (Roche)

NOTE

Please ensure that all instruments used have been installed, calibrated, checked and maintained according to the manufacturer's instructions and recommendations.

5. Procedure

5.1 Sample Preparation

Extracted DNA is the starting material for the RealStar® JCV PCR Kit 1.2.

The quality of the extracted DNA has a profound impact on the performance of the entire test system. It has to be ensured that the system used for nucleic acid extraction is compatible with real-time PCR technology. The following kits and systems are suitable for nucleic acid extraction:

- QIAamp® DNA Mini Kit (QIAGEN)
- QIAamp® Viral RNA Mini Kit (QIAGEN)
- QIASymphony® (QIAGEN)
- NucliSENS® easyMag® (bioMérieux)
- MagNA Pure 96 System (Roche)
- m2000sp (Abbott)
- Maxwell® 16 IVD Instrument (Promega)
- VERSANT® kPCR Molecular System SP (Siemens Healthcare)

Alternative nucleic acid extraction systems and kits might also be appropriate.

If using a spin column based sample preparation procedure including washing buffers containing ethanol, it is highly recommended to perform an additional centrifugation step for 10 min at approximately 17000 x g (~ 13000 rpm), using a new collection tube, prior to the elution of the nucleic acid.

CAUTION

If your sample preparation system is using washing buffers containing ethanol, make sure to eliminate any traces of ethanol prior to elution of the nucleic acid. Ethanol is a strong inhibitor of real-time PCR.



The use of carrier RNA is crucial for extraction efficiency and stability of the extracted nucleic acid.

For additional information and technical support regarding pre-treatment and sample preparation please contact our Technical Support (see chapter 8. Technical Assistance).

5.2 Master Mix Setup

All reagents and samples should be thawed completely, mixed (by pipetting or gentle vortexing) and centrifuged briefly before use.

The RealStar® JCV PCR Kit 1.2 contains a heterologous Internal Control (IC), which can either be used as a PCR inhibition control or as a control of the sample preparation procedure (nucleic acid extraction) and as a PCR inhibition control.

- ▶ If the IC is used as a PCR inhibition control, but not as a control for the sample preparation procedure, set up the Master Mix according to the following pipetting scheme:

Number of Reactions (rxns)	1	12
Master A	5 µl	60 µl
Master B	10 µl	120 µl
Internal Control	1 µl	12 µl
Volume Master Mix	16 µl	192 µl

- ▶ If the IC is used as a control for the sample preparation procedure and as a

PCR inhibition control, add the IC during the nucleic acid extraction procedure.

- ▶ No matter which method/system is used for nucleic acid extraction, the IC **must not** be added directly to the sample. The IC should always be added to the sample/lysis buffer mixture. The volume of the IC which has to be added, always and only depends on the elution volume. It represents 10% of the elution volume. For instance, if the nucleic acid is going to be eluted in 60 µl of elution buffer or water, 6 µl of IC per sample must be added into the sample/lysis buffer mixture.
- ▶ If the IC was added during the sample preparation procedure, set up the Master Mix according to the following pipetting scheme:

Number of Reactions (rxns)	1	12
Master A	5 µl	60 µl
Master B	10 µl	120 µl
Volume Master Mix	15 µl	180 µl

CAUTION



If the IC (Internal Control) was added during the sample preparation procedure, at least the negative control must include the IC.



No matter which method/system is used for nucleic acid extraction, never add the IC directly to the sample.

5.3 Reaction Setup

- ▶ Pipette 15 µl of the Master Mix into each required LightCycler® capillary or reaction tube for the SmartCycler® II.
- ▶ Add 10 µl of the sample (eluate from the nucleic acid extraction) or 10 µl of the controls (Quantification Standard, Positive or Negative Control).

Reaction Setup	
Master Mix	15 µl
Sample or Control	10 µl
Total Volume	25 µl

- ▶ Make sure that at least one Positive (QS) and one Negative Control is used per run.
- ▶ For quantification purposes all Quantification Standards (QS1 to QS4) should be used.
- ▶ Thoroughly mix the samples and controls with the Master Mix by pipetting up and down.
- ▶ Close the capillaries or the tubes using appropriate lids.
- ▶ Centrifuge the LightCycler® capillaries or reaction tubes of the SmartCycler® II using an appropriate centrifuge for 30 seconds at approximately 400 x g (~2000 rpm).

6. Programming the Real-Time PCR Instrument

For basic information regarding the setup and programming of the different real-time PCR instruments, please refer to the user manual of the respective instrument.

For detailed programming instructions regarding the use of the RealStar® JCV PCR Kit 1.2 on specific real-time PCR instruments please contact our Technical Support (see chapter 8. Technical Assistance).

6.1 Settings

- Define the following settings:

Settings	
Reaction Volume	25 µl*
Ramp Rate	Default

* The reaction volume has to be defined as 20 µl, if using a LightCycler® 2.0 Instrument (Roche).

6.2 Fluorescence Detectors (Dyes)

- Define the fluorescence detectors (dyes):

Target	LightCycler® 1.2/1.5	LightCycler® 2.0	SmartCycler® II
JCV specific DNA	F1	530	FAM™
Internal Control (IC)	F2	610	Cy®3

CAUTION

For accurate data analysis on the LightCycler® Instruments a specific Color Compensation File might be needed. Please contact altona Diagnostics GmbH for assistance



If using the LightCycler® 2.0 Instrument, only the detection channels 530 and 610 should be activated for color compensation.

6.3 Temperature Profile and Dye Acquisition

- Define the temperature profile and dye acquisition:

	Analysis Mode	Cycle Repeats	Acquisition	Temperature [°C]	Time [min:sec]
Denaturation	None	1	-	95	02:00
Amplification	Quantification	45	None	95	00:05
			Single	60	00:30
			None	72	00:10
Cooling	None	1	-	40	00:30

7. Data Analysis

For basic information regarding data analysis on specific real-time PCR instruments, please refer to the user manual of the respective instrument.

For detailed instructions regarding the analysis of the data generated with the RealStar® JCV PCR Kit 1.2 on different real-time PCR instruments please contact our Technical Support (see chapter 8. Technical Assistance).

7.1 Interpretation of Results

7.1.1 Qualitative Analysis

Detection Channel		Result Interpretation
FAM™/F1/530	Cy®3/F2/610	
+	+*	JCV specific DNA detected.
-	+	No JCV specific DNA detected. Sample does not contain detectable amounts of JCV specific DNA.
-	-	PCR inhibition or reagent failure. Repeat testing from original sample or collect and test a new sample.

* Detection of the Internal Control in the Cy®3/F2/610 detection channel is not required for positive results in the FAM™/F1/530 detection channel. A high JCV DNA load in the sample can lead to a reduced or absent Internal Control signal.

7.1.2 Quantitative Analysis

The RealStar® JCV PCR Kit 1.2 includes four Quantification Standards (QS). In order to generate a **standard curve** for quantitative analysis, these have to be defined as **standards** with appropriate concentrations (see chapter 4. Product Description). Using **standards** of known concentrations a standard curve for quantitative analysis can be generated.

$$C_t = m \cdot \log(N_0) + b$$

C_t = Threshold Cycle

m = Slope

N_0 = Initial Concentration

b = Intercept

Derived from the standard curve positive samples of unknown concentrations can be quantified.

$$N_0 = 10^{(C_t - b) / m}$$

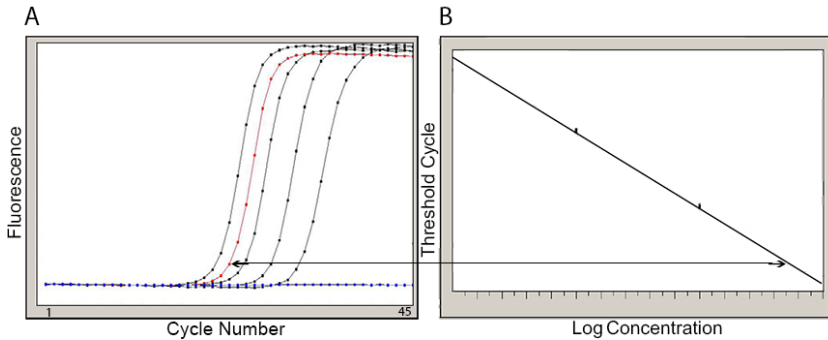


Figure 1: Quantification Standards (black), a positive (red) and a negative sample (blue) displayed in the Amplification Plot [A] and Standard Curve analysis [B]

NOTE



The concentration of the "Sample" is displayed in IU/μl and refers to the concentration in the eluate.

To determine the **viral load of the original sample**, the following formula has to be applied:

$$\text{Viral load (Sample) [IU/ml]} = \frac{\text{Volume (Eluate) } [\mu\text{l}] \cdot \text{Viral load (Eluate) [IU/\mu\text{l}]}{\text{Sample Input [ml]}}$$

8. Technical Assistance

For technical advice, please contact our Technical Support:

e-mail: **support@altona-diagnostics.com**

phone: **+49-(0)40-5480676-0**

9. Trademarks and Disclaimers
















RealStar® (altona Diagnostics); ABI Prism® (Applied Biosystems); ATCC® (American Type Culture Collection); CFX96™ (Bio-Rad); Cy® (GE Healthcare); FAM™, JOE™, ROX™ (Life Technologies); LightCycler® (Roche); SmartCycler® (Cepheid); Maxwell® (Promega); Mx 3005P™ (Stratagene); NucliSENS®, easyMag® (bioMérieux); Rotor-Gene®, QIAamp®, MinElute®, QIASymphony® (QIAGEN); VERSANT® (Siemens Healthcare).

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For research use only (RUO)! Not for use in diagnostic procedures.

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10. Explanation of Symbols

Symbol	Explanation
	For Research Use Only
	Batch code
	Cap color
	Product number
	Content
	Number
	Component
	Consult instructions for use
	Contains sufficient for “n” tests/reactions (rxns)
	Temperature limit
	Use-by date
	Manufacturer
	Caution
	Note
	Version

Notes:

always a drop ahead.

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